EFFECT OF DIFFERENT DIETS ON SURVIVAL AND GROWTH OF STRIPED MURREL FRY CHANNA STRIATUS

D. Kumar & M.A. Haniffa

ABSTRACT
Tubifex, Chironomus, Beef liver, mosquito larvae and plankton used as feed were tested as diets in the early larval growth of Channa striatus fry over a 45 days period. Among all the food types Chironomus was found to produce the best growth results followed by tubifex, while other food types yielded poor results.

Key words: Channa striatus, growth, nutrition, diet

Introduction
Various dry feed formulae have been investigated as possible substitutes of live food for larval development (Appelbaum and Dor, 1978; Dabrowski 1983; 1984). In recent years suitability of various dry feed formulae has been investigated for the rearing of Cyprinid and catfish larvae (Bryant and Matty 1981; Msiska, 1981; Hecht and Viljoen, 1982). However it has been shown that formulated compound diets do not provide optimal larval growth when used exclusively as larval food, especially during the early larvae stages of Cyprinids and catfish (Hogendoorn, 1980; Dabrowski 1984; prinlsoo and Schoonbee, 1986), therefore live food provide

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a substantial availability of protein and other essential nutrients (Jhingran, 1975; Ahmed, 1994; Thakur, 1978; Munnet, 1979) provided some basic information on the feeding of C. batrachus fry. However development of suitable feed for rearing Channa striatus fry is lacking. It is therefore important to study the efficacy of a few selective feeds.

Channa striatus snakehead is an air breathing, and carnivorous in nature. They are widely distributed in Africa and Asia. It is having high market due to its taste and flesh. They support economically important fisheries and aquaculture industry in many Asian countries (Ling 1979, Chen 1990). Among murrels C. striatus forms a significant role in capture fisheries of India. Characteristics of this fish that make it desirable cultivable food fish include rapid growth and the ability of the fish to store and use atmospheric oxygen for respiration in waters with low dissolved oxygen and they can withstand higher stocking densities also. It has been estimated that out of 18,000 t of marketable surplus air breathing fishes caught from natural resources in India, murrels account for nearly 12,000 t (Jhingran 1975) with major part of them constituted by Channa marulius, Channa striatus and Channa punctatus. However, murrel culture is not practised in a well-defined way in India due to several reasons. One of these is that there are no seed supply / sales centre for murrels in country.

The fish farmers therefore depend on wild collection, which are unpredictable. Further, the rearing of hatchlings, post larvae and fry of C. striatus is a complicated process unlike the raising of carp fry, which has been standardized in some extent. Recently attempts have been made on larval nutrition of Channa striatus by Qin et al. (1997), Samantaray and Mohanty (1997). But these authors have provided formulated pelleted feeds (instead of live feed) to the larvae resulting in poor survival and growth. In the present study a comparison is made on the growth of C. striatus larvae using tubific, Chironomus, plankton and Beef liver and mosquito larvae as two alternative live foods during the early larval growth phase of this species.

**Materials and Method**

Channa striatus fry (length: 3.57 + 0.05 cm, weight 0.425 + 0.03 g) were collected from (CARE) earthen pond and acclimatized in the cement tanks for a period of one week; during this period they were fed with plankton soup. They were grouped into 5 batches and stocked for 10 individual for each treatment with three replicates and reared in plastic troughs. (Capacity 15/Lt).

Water quality parameters viz., temperature 29°C ± 1°C dissolved oxygen 6.1-6.6 mg/Lt and pH 7.5-8.25 were recorded throughout the study. They were fed with
tubifex, Chironomus larvae, beef liver, mosquito larvae and plankton soup twice/day (11.00 hrs, 13.00 hrs) ad libitum. The feeding trial was continued for a period of 45 days. Water was changed everyday with minimal disturbance to the experimental animals full nos. the length and weight were recorded once in every fortnight, the growth parameters viz, weight gain (%), specific growth rate and (%/day) and survival (%) were estimated as Weight gain (%/day)=final mean weight−initial mean weight / initial mean weight / days x 100; SGR (%/day)=final mean log weight−initial mean log weight/days x 100; Survival (%) = Final total no. of fish/Initial total no. of fish x 100. After the completion of the experiment five fishes were sacrificed for the proximate composition estimations.

Results

All foods were readily accepted from the start of feeding. The beef liver was taken by larvae from the bottom of the trough, whereas zooplankton, mosquito larvae in the mid water of the trough and the tubifex, Chironomus taken by the larvae from the bottom. The body composition of the fish fed on different diets and growth parameter was presented in table 1, 2. From that tables, body protein, carbohydrate and lipid of the fish is higher in the diet of D2 and followed by other diets groups. And Fig 1 shows the Specific growth rate and relative growth rate and growth rate of Channa striatus fry.

Table 1. Growth and Survival of Channa striatus fry fed on different types of diets

<table>
<thead>
<tr>
<th></th>
<th>Tubifex</th>
<th>Blood worm</th>
<th>Beef liver</th>
<th>Mosquito</th>
<th>Plankton</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial length (cm)</td>
<td>3.506±.024</td>
<td>3.603±.018</td>
<td>3.586±.109</td>
<td>3.66±.141</td>
<td>3.523±.053</td>
</tr>
<tr>
<td>Initial weight (g)</td>
<td>.406±.020</td>
<td>.390±.031</td>
<td>.425±.041</td>
<td>.443±.040</td>
<td>.463±.026</td>
</tr>
<tr>
<td>Final length (cm)</td>
<td>6.918±.072</td>
<td>7.66±.082</td>
<td>6.69±.052</td>
<td>6.512±.305</td>
<td>6.054±.069</td>
</tr>
<tr>
<td>Final weight (g)</td>
<td>3.267±.020</td>
<td>3.948±.056</td>
<td>2.85±.081</td>
<td>2.11±.082</td>
<td>2.126±.010</td>
</tr>
<tr>
<td>Duration</td>
<td>45</td>
<td>45</td>
<td>45</td>
<td>45</td>
<td>45</td>
</tr>
<tr>
<td>SGR%</td>
<td>1.383±.049</td>
<td>1.529±.052</td>
<td>1.22±.097</td>
<td>1.112±.081</td>
<td>1.082±.045</td>
</tr>
<tr>
<td>Weight gain</td>
<td>705.64±</td>
<td>919.00±</td>
<td>578.15±</td>
<td>380.20±</td>
<td>360.185±</td>
</tr>
<tr>
<td></td>
<td>41.805</td>
<td>62.324</td>
<td>74.78</td>
<td>38.37</td>
<td>26.47</td>
</tr>
<tr>
<td>RGR%</td>
<td>7.056±.418</td>
<td>9.162±.70</td>
<td>5.78±.747</td>
<td>3.801±.382</td>
<td>3.601±.264</td>
</tr>
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</tr>
<tr>
<td>ADG (g/day)</td>
<td>.0723±.0004</td>
<td>.0674±.0013</td>
<td>.0629±.0016</td>
<td>.0466±.001</td>
<td>.0472±.0002</td>
</tr>
<tr>
<td>Growth rate (%)</td>
<td>39.39±.301</td>
<td>40.55±.299</td>
<td>37.79±.600</td>
<td>35.56±.760</td>
<td>35.18±.551</td>
</tr>
<tr>
<td>Survival%</td>
<td>90±14.14</td>
<td>96.66±4.714</td>
<td>76.66±4.714</td>
<td>66.66±4.714</td>
<td>73.33±9.428</td>
</tr>
</tbody>
</table>

Table 2. Body composition of the Channa striatus fry fed on different diets

<table>
<thead>
<tr>
<th></th>
<th>Initial</th>
<th>D1</th>
<th>D2</th>
<th>D3</th>
<th>D4</th>
<th>D5</th>
</tr>
</thead>
<tbody>
<tr>
<td>Protein %</td>
<td>14.01</td>
<td>15.14</td>
<td>16.28</td>
<td>15.68</td>
<td>14.21</td>
<td>14.32</td>
</tr>
<tr>
<td>Carbohydrate %</td>
<td>.88</td>
<td>.98</td>
<td>1.01</td>
<td>.88</td>
<td>.94</td>
<td>.93</td>
</tr>
<tr>
<td>Lipid %</td>
<td>3.14</td>
<td>3.16</td>
<td>3.06</td>
<td>2.94</td>
<td>3.01</td>
<td>2.84</td>
</tr>
<tr>
<td>Ash %</td>
<td>3.2</td>
<td>3.5</td>
<td>3.9</td>
<td>2.8</td>
<td>3.2</td>
<td>3.4</td>
</tr>
<tr>
<td>Moisture%</td>
<td>76.14</td>
<td>74.15</td>
<td>73.48</td>
<td>75.14</td>
<td>74.28</td>
<td>75.01</td>
</tr>
</tbody>
</table>

D1-Tubifex, D2-Chironomus, D3-Beef liver, D4-Mosquito, D5-Plankton

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**Fig 1.** Effect of different diets on the growth performance of Channa striatus fry fed on different diets

Live food is regarded as the best for fishes, many crustacean, insect larvae. In present study the result revealed that the highest specific growth rate (1.529%/day), was obtained in the individual fed with Chironomus (Table 1) followed by tubifex fed on 1.383%/day), beef liver (1.22%/day), mosquito larvae (1.112%/day) and plankton (1.082%/day). The best weight gain was obtained by those fed on
Chironomus larvae (919.00). And followed by (705.64) those fed on tubifex. The best survival rate (96.66 %) was obtained in Channa striatus fed on Chironomus larvae whereas the lowest survival (73.33%) were observed in those fed on plankton.

Discussion

The results showed that live food, in particular Chironomus larvae is a most desirable diet for the rearing of the Channa striatus larvae. The importance of artemia as live food (Hogendoorn, 1980; Msiska, 1981) is again confined by this investigation. However Prinsloo and Schoonbee (1986) observed zooplankton as best live food in comparison to commercial dry feed for the rearing of the silver carp and grass carp species over a period of 10-14 days; silver carp and grass carp larvae acquired relatively better growth with zooplankton as compared to commercial dry food. In our present study instead of zooplankton chironomus displayed superior growth and other parameter were recorded, due to the higher protein 61.17% and haem content of the Chironomus larvae. Qin, Fast, De Anda & Weidenbach (1997) also developed a protocol for weaning larval snake heads from live artemia to formulated feed, but grow out performance with formulated feed was not evaluated.

Live food is an important diet in the rearing of larvae of a member of fish species (Hogendoorn, 1980; Msiska, 1981; Stenson; 1982) indicated the importance of rotifer Brachionus plicatilis for mass larval rearing of fishes and stressed the value of the inclusion of rotifers in combination with artificial dry feeds for the optimum growth of Cyprinus carpio larvae. Matlik and Matlak (1976) indicated that rotifers are important food items of carp larvae during the first three weeks in nursery ponds. Zooplankton is the best larval food for a variety of fish larvae (Kilambi and Zdinak, 1982; Geiger 1983a, and Dabrowski 1984). The nutritional value of artemia for Cyprinus carpio larvae indicated good growth (Bryant and Matty 1981). A variety of dry foods were used for the rearing of C.carpio larvae (Appelbaum and Dor 1978; Hecht and Viljean 1982).

According to Cahu et al. (1998), larvae receiving live food also showed better survival and growth than larvae receiving artificial diets, sea bass Dicentrarchus labrax. Abi-Ayad and Kestemont (1994) observed the highest SGR in gold fish Carassius auratus larvae fed the mixed diets. Whereas the lowest SGR (1.325%/day) fed on plankton. Numbers of reports are available on larval rearing, Mahseer larvae (Rai, 1990), also reported that larvae of Mahseer fed with plankton showed better growth when fed with other supplementary diets. In addition to the fry of kattle Acrossocheilus hexagonolepis also showed better growth, when fed with 30% protein content with plankton soup (Rai, 1990).
Malkota and Munch (1985) found that formulated feed might also be physically unsuitable for most first feeding fish larvae because large food particles that didn’t pass down the gut could subject larvae to physical stress or physiological stress. The fish growth rate is generally related to availability and density of optimal food (Mittelbach 1981). Walleye growth increased after they switched their diets from zooplankton to chironomids (Fox et al 1989). But in our present study Chironomus larvae displayed superior growth fed with the Channa striatus fry is reported.

Dabrowski (1982) reported that many small fish larvae do not have the enzymes for digesting non-living diets. So we are applied different types of feeds. Artemia nauplia suitable for rearing of the young ones. Some relevant reports are available; Fluechter (1980) found that protein digestion enzymes in live Artemia nauplia were responsible for successful rearing of White fish (coregonus lavaretus) larvae.

Conclusion

Our present results strongly support the use of live food in the early larval growth phase of C.striatus larvae. So the blood worm is strongly recommended to the rearing of C.striatus larvae, it should not affect further development of the larvae.

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References


